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Molecular Recognition of cis-1,3,5-Cyclohexane Tricarboxylic Acid

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Abstract: The **design and synthesis of a new receptor designed to bind tricarboxylic acids in organic** solvents is described. The properties of the complex formed between the new receptor and cis-1,3,5cyclohexanetricarboxylic acid are studied.

Molecular recognition of carboxylic acids in organic solvents has been traditionally achieved by the use of neutral receptors with complementary hydrogen bonding groups¹⁻³. For example, Hamilton² and Diederich³ have both synthesized several neutral receptors for the selective binding of dicarboxylic acids in organic solvents by separating two amidopyridine residues with a suitable spacer. The 2-amidopyridine group can form two hydrogen bonds, one to the acidic proton and one to the syn lone pair of the carboxylic acid carbonyl4. We rationalized that if a relatively large and rigid molecular component was used to adequately position three 2 amidopyridines groups, then a potential receptor for the formation of stable host-guest complexes with tricarboxylic acids in organic solvents would be at hand.

Our recent experience⁵ in the use of the tetrachlorosilane-ethanol induced self condensation of ketones⁶ for preparing 1,3,5-tris(3-bromo-5-carboxyphenyl-4-hydroxy-}benzene, prompted us to consider the use of compound **2a** (m-p. 222-225°C) as a rigid molecular component for **positioning** three hydrogen bonding arrays within a receptor. Molecular modeling studies7 on the newly designed receptor **3a suggested** that the syn conformation of the molecule was very well suited for the complexation of cis-1,3,5-cyclohexanetricarboxylic acid 4 (Figure 1). In this conformation the hydrogen bonding groups converge and can act simultaneously in "grasping" the guest molecule in the induced cleft of the receptor.

It is worth mentioning that we made use of intramolecular hydrogen bonding between the amide carbonyls and the phenolic protons for controlling receptor preorganization.⁸ The other possible conformer in which the

amide protons are hydrogen-bonded to the phenolic oxygens is expected to be largely disfavored⁹. Thus, the favored intramolecular hydrogen bonds restrict the rotation of the $C_{\text{aryl}}-C_{\text{amide}}$ bonds forcing the amide hydrogens to be directed towards the interior of the cavity and locking each amidopyridine subunit in an almost planar configuration with respect to their aryl ring. Receptor 3a (m.p. > 310°C) was synthesized uneventfully in five steps from 4-acetylmethylsalicylate 1 in 31% overall yield¹⁰.

Unfortunately, host 3a was insoluble in CHCl3 or other organic solvents which do not considerably disrupt hydrogen bonding. in order to increase the solubility of the receptor in CHC13, a more lipophilic version of the spacer was prepared which involved etherification of the phenols of $2a$ with allyl bromide¹¹, followed by a triple Claisen¹² rearrangement and catalytic hydrogenation of the double bonds to afford the propylated triaryl benzene triester 2b (m.p. 202-205°C, 61% overall yield). Triamide 3b (m.p. 281-284°C) was prepared from 2b by a slight modification of the Weinreb¹³ procedure in 26% yield (Scheme)¹⁴. Chloroform solutions of receptor 3b of up to 10^{-2} M could now be prepared.

Scheme. (a) $Cl_4Si/EtOH$, r.t., 24h; (b) Cs_2CO_3 , DMF/acetone, Br-CH₂-CH=CH₂ reflux, 24h; (c) N,N-dimethylaniline, reflux, 12h; (d) H_2 , Pd/C 10%, ethyl acetate, 10 psig, 5h; (e) 3 equiv Me₃Al, then Me₃Al/2-amino-6-methylpyridine complex, benzene reflux, 48h.

The IH-NMR **spectrum of** 3b **in** CDC13 showed that the phenolic protons are shifted downfield to 12.4 ppm indicating that they are strongly hydrogen bonded while the amide protons appears at 8.8 ppm, a reasonable position for free aromatic amide protons, These observations strongly support the confotmational preference aforementioned. At room temperature, Caryl-Caryl bond rotations are fast in the NMR time scale and the signals are sharp and well resolved. At low temperature, broadened signals are observed for 3b since interconversions between different conformers become slower.

Addition of solid cis-1,3,5-cyclohexanetricarboxylic acid 4 to a CDC13 solution of 3b led to a rapid dissolution of the normally insoluble substrate. Large downfield shifts (2.8 ppm) of the NH resonances in the $1H\text{-NMR spectrum indicated the formation of a hydrogen-bonded complex, which integration established as a$ 1: 1 stoichiometry. The spectrum of the 1: 1 complex formed by slid-liquid extraction was no longer temperature dependent, Thus, the binding of the triacid 4 restricts the conformation of **3b** to the one featuring convergent hydrogen bonding. Only this conformation can involve all three acylamidopyridine groups of the receptor, thus freezing the three Caryl-Caryl bond rotations. Host 3b and triacid 4 crystallize out of a chloroform/cyclohexane solution as a 1:1 complex, as shown by 1 H-NMR. Attempts to obtain an x-ray crystal structure have not yet been successful, although single crystals have been obtained.15

Molecular modeling studies suggest two e3-symmetric geometries for a complex between 3b and 4 in which six intermolecular hydrogen bonds between the three COOH residues of 4 and the three CONH(py)N groups of **3b** are possible (Figure 2). In geometry A, the acids and He hydrogens of the guest are endo to the receptor cavity, while in geometry B , the acids are exo and the H_d hydrogens are endo. Both geometries were

optimized with MacroModel 3.5X using **the OPLS *16** force field and the GBISA-CHCl3 solvation model", which resulted in an energy difference of $\Delta E = 8$ kcal/mol in favor of binding geometry A. The calculated energy difference could be attributed **to the fact that** intermolecular hydrogen bonding in complex geometry **B** may not be optimal. In fact, modeling suggests that the three (py)N₁₁. H-OOC hydrogen bonds in geometry B are longer (3.01Å vs. 2.91Å) and less collinear (150 $^{\circ}$ vs. 174 $^{\circ}$) than in geometry A, which may account for the calculated energy difference.

Figure 2. Above, two different geometries of the convergent complex between triacid 4 and receptor 3b. Below, schematic representations of the selected areas highlighting bond lenghts and bond angles for the hydrogen bonds in the complexes. $R = propyl$.

The A **type** binding geometry was confirmed experimentally by 2D IH-NMR studies of the 1 **:l complex** in CDCl₃ solution. A ROESY experiment on the 1:1 complex (mixing time=0.75 s., spinlock=2KHz) revealed close intermolecular contacts between the H_a and H_b hydrogens of the receptor and the H_c hydrogens of the triacid. This observation, together with the absence of rGe's between the receptor and the triacid's Ha hydrogens, provides strong evidence for the predominance of the type A complex geometry in chloroform solution.

The association constant between triacid 4 and triamide 3b was estimated by the solubility titration method¹⁸. By plotting the insoluble component 4 vs. soluble component **3b** and by simple analysis of the slope and intercept, we determined the association constant for the complex $K_a > 10^5$ L mol⁻¹. Furthermore, a standard solution NMR titration experiment,¹⁹ made possible by using 20%d⁸-THF/CDCl₃ as solvent, gave also an association constant $K_a > 10^5 L$ mol⁻¹.

To the best of our knowledge, this is the first report of an organic soluble receptor for a triacid. The binding geometry of the complex between **3b** and 4 has been probed by molecular modeiing and 2D nGe experiments. The high association constants observed for the complex reflect a precise match between receptor and guest in establishing a panoply of intermolecular interactions. Studies on the binding selectivity of receptor **3b** to several other triacids are in progress and will be reported in due time.

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